Custom Sigma CRISPRs Application Form

Instructions

The information in this form will be used by Sigma to generate a quotation for your particular project and to anticipate any potential risks in its production and your application. **Please answer all questions in sections 1 to 6**, and then answer section 7 for Gene Knockout Projects, section 8 for Gene Modification Projects, or section 9 for Targeted Integration Projects. Sections 12–15 are for donor design. Sections 10 and 11 are optional.

After completing the form, please email it to **CRISPR@sial.com**. Once we receive your order, a CRISPR design and production specialist will contact you to discuss and agree upon a detailed CRISPRs design strategy.

1. Name	1a. Title	1b. First Name	1c. Last Name
2. Address	2a. Institution or company name		
	SHIPPING ADDRESS		
	2b. Building and street address		
	2c. City or town, state or province, and country. Include ZIP code or postal code where appropriate.		
3. Contact Details	3a. Email address	3b. Telephone. Include country and area code.	
4. Project Goals	4a. Please indicate the type of project. ☐ Gene Knockout ☐ Targeted Gene Integration ☐ Gene Modification ☐ Other (please describe) _____		
	4b. What is the final format of your project? ☐ Cell line and/or ☐ Whole animal		
	4c. What is your intended application? ☐ Basic Research ☐ Target ID / Validation ☐ Screening ☐ Bioproduction ☐ Model cell line / animal ☐ Other, please describe _____		
	4d. Is there an intended commercial application for this project? ☐ No ☐ Bioproduction ☐ Food Production ☐ Textile Production ☐ Companion Animal ☐ Other, please describe_____		
	4e. Are you interested in learning more about CompoZr ZFNs, the gold standard in genome editing? ☐ Yes ☐ No		
5. CRISPR Format	5. Indicate the format you would like to receive your Sigma CRISPRs in:		
	5a. Plasmid – check all that apply: All-in-One Vector gRNA + Cas9 WT – plasmid: ☐ GFP or ☐ RFP Dual Vector Cas9 WT: ☐ gRNA – plasmid ☐ Cas9 WT plasmid Paired Cas9 Nickase: ☐ Cas9 nickase plasmid ☐ Paired gRNA plasmid		
	5b. mRNA – check all that apply: Cas9 WT: ☐ Cas9 WT mRNA ☐ gRNA – RNA only Paired Cas9 Nickase: ☐ Cas9 nickase mRNA ☐ Paired gRNA – RNA only		

6. Target Gene(s)	6a. What organism's genome are you interested in editing? (please select one) ☐ Human ☐ Mouse ☐ Rat ☐ Zebrafish ☐ Other (*genus species*): _____		
	IF ORDERING FOR 3+ GENES PLEASE ATTACH SPREADSHEET WITH THE FOLLOWING INFORMATION:		
	6b. Name of target gene(s):	6c. Target gene Id(s):	6d. mRNA accession number (eg, NM_000111):
	6e. For a target gene with multiple transcript variant(s), please specify which isoforms would you like to target (mRNA accession number required). mRNA accession number_____		
	6f. Are there any pseudogenes or homologs for your target? If yes, which ones should be targeted? ☐ Yes, please target: _____		
	6h. How many copies are in your cell line or organism?	6i. Is it essential to cell viability or organism development? ☐ Yes ☐ No	
7. Gene Knockout Project	7a. Is standard targeting within the first 2/3 ORF acceptable to your application? ☐ Yes ☐ No		
	7b. If 'No', please specify the target region(s) and provide its or their DNA sequence(s) for CRISPR design in detail below:		
8. Gene Modification Project	8. Please attach target sequence to be modified, either as cDNA sequence or at least 200 nt of genomic sequence, with specified mutation position(s) marked.		
9. Targeted Integration Projects	9a. Do you wish to maintain function of the gene product when modifying the allele? ☐ Yes ☐ No		
	9b. Please select and specify integration site(s) on the target gene: ☐ At N-terminus of the target gene. Typically 200 nt centering around the start codon will be used for CRISPR design. ☐ At C-terminus of the target gene. Typically 200 nt centering around the stop codon will be used for CRISPR design. ☐ At either N-terminus or C-terminus of the target gene. ☐ Other integration regions. Please specify the regions for tagging and provide the DNA sequences for CRISPR design.		

10. Other Design Information	10. Are there any other details about your Gene-Editing objective that you would like to share with us in order to design and deliver the most effective CRISPRs for your project goal? Please provide details here.	
11. Cell Design Studio (CDS)	11a. Do you want CDS to engineer your cell line? ☐ Yes ☐ No	11b. Is this cell line available commercially? ☐ Yes ☐ No

THIS SECTION FOR DONOR DESIGN REQUESTS ONLY

We strongly encourage identifying active CRISPRs BEFORE designing donors.
Additional charges apply for donor design.

12. Project Information	12a. Model system: ☐ Cells ☐ Embyo injection	12b. Preferred donor format: ☐ dsDNA ☐ ssDNA oligo
13. Insertion Projects	13a. Please indicate the insertion position(s) in your sequence of interest. Include ≥50 bases on each side and denote the insertion position with an underscore (e.g.,"ACTG_ACTG").	
	List all exogenous elements to be inserted. Begin with the most 5′ element; end with the most 3′ element. (example: i. CMV promoter, ii. GFP, iii. polyA…)	
	13b. Name	13c. Sequence (if available)
	i.	
	ii.	
	iii.	
	iv.	
	v.	
	vi.	
	vii.	
14. Modification or Deletion Projects	Nucleotide sequence of locus before targeting, and desired nucleotide sequence after targeting (include ≥ 50 bases of sequence flanking the modification or deletion site(s)):	
	14a. Sequence before:	14b. Sequence after:
15. Donor Modifications	15. RFLP mutations? ☐ Yes ☐ No	

Internal use only: